This is the published version of a paper published in *Journal of Oral Microbiology*.

Citation for the original published paper (version of record):

Detection of a 640-bp deletion in the Aggregatibacter actinomycetemcomitans leukotoxin promoter region in isolates from an adolescent of Ethiopian origin.
*Journal of Oral Microbiology*, 7: 26974
http://dx.doi.org/10.3402/jom.v7.26974

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NOTE

Detection of a 640-bp deletion in the Aggregatibacter actinomycetemcomitans leukotoxin promoter region in isolates from an adolescent of Ethiopian origin

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The expression of the leukotoxin of Aggregatibacter actinomycetemcomitans is regulated by the leukotoxin promoter. A 530-bp deletion or an 886-bp insertion sequence (IS) element in this region has earlier been described in highly leukotoxic isolates. Here, we report on highly leukotoxic isolate with a 640-bp deletion, which was detected in an adolescent of Ethiopian origin.

Keywords: leukotoxicity; deletion; JP2; aggressive periodontitis; geographic spreading; Africa

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Received: 14 December 2014; Revised: 3 March 2015; Accepted: 10 March 2015; Published: 13 April 2015

The Gram-negative oral bacterial species, Aggregatibacter actinomycetemcomitans, expresses a leukotoxin that links the bacterium to aggressive periodontitis (1–6). The toxin expression, which is associated with four structural genes (ltxA, ltxB, ltxC, and ltxD), is regulated by the leukotoxin promoter (Fig. 1). Although the genes of the leukotoxin operon appear to be present in all strains of A. actinomycetemcomitans, the toxin expression varies substantially between different strains (7–9). High expression of leukotoxin has been detected in A. actinomycetemcomitans isolates with a 530-bp deletion in the promoter region and in isolates carrying an 886-bp IS element in the same region (7–10) (Fig. 1).

Isolates characterized by the 530-bp deletion comprise the so-called JP2 genotype, and isolates with a complete leukotoxin promoter, which consequently are distributed within the group often called the non-JP2 genotype, are described as low or moderately leukotoxic (11). Reports about other leukotoxin promoter types than the JP2, the non-JP2 genotype, and the genotype with the IS element are absent.

The JP2 genotype of A. actinomycetemcomitans, which has been studied for more than 20 years from an experimental perspective, is almost exclusively isolated from individuals of African origin living in North and West Africa (12, 13). However, a Sudanese subject, positive for the JP2 genotype, was reported recently (14). Although much rarer, findings of the JP2 genotype among non-Africans also have been described, indicating that this genotype is more widespread than was initially believed (15–17).

Based on longitudinal studies, individuals carrying the JP2 genotype have an enhanced risk of developing aggressive periodontitis (3, 5, 6). However, an association with aggressive periodontitis among carriers of the non-JP2 genotype cannot be neglected (6, 18, 19).

Recently, we identified a leukotoxin promoter type of A. actinomycetemcomitans, which has not previously been reported. It was detected in two periodontal plaque samples collected from a 16-yr old female periodontitis patient of Ethiopian descent (Fig. 2). The samples were sent in vials containing VMGA III medium (20) to our clinical laboratory at the Dental School in Umeå, Sweden. After a procedure as described (17), it was found that the plaque samples contained high proportions of A. actinomycetemcomitans (Table 1). According to the routine procedure, two isolates (456A1, 456A2) were serotyped according to Suzuki and co-workers (21). Since the two isolates belonged to serotype b, they were typed according to the leukotoxin promoter as described (22). Surprisingly, both isolates were...
characterized by a leukotoxin promoter region that was approximately 100 bp smaller than the JP2 promoter region, i.e., they harboured a previously undescribed promoter type (Figs. 1 and 2). This finding was confirmed when the bacterial DNA, taken directly from the samples (455V and 456V) in the transport vials (VMGA), was amplified with leukotoxin promoter-specific primers (Fig. 2). The identical characteristics of the analyzed isolates and samples indicate that the patient most likely is colonized by a single clone of *A. actinomycetemcomitans*.

DNA was purified from one of the two isolates (456A1) with a DNA-purifying kit (Ready-To-Go Kit; GE Healthcare, Little Chalfont, UK) for further characterization. First, when the promoter region was sequenced as described (6), it was found to be associated with a 640-bp deletion, i.e., it lacked 640 bp as compared with strains with the full-length leukotoxin promoter (Fig. 1). In comparison with the 530-bp deletion in the leukotoxin promoter region in JP2 genotype strains, the additional 110 bp missing in the sequenced isolate (456A1) were located 108 bp upstream and 2 bp downstream from the site of the JP2 genotype/C1 associated deletion. Although isolate 456A1 carried a different leukotoxin promoter than JP2 genotype strains, it was found to have leukotoxicity similar to that of the JP2 genotype of *A. actinomycetemcomitans* (strain HK1615), when leukotoxic activity was determined as described (6) (Fig. 3).

By sequencing the hemoglobin binding gene (*hbp2*), JP2 genotype isolates from North Africa, which have an ‘A’ nucleotide in position 525285 (213 within the 329-bp gene), are distinguished from isolates of West African origin, which have a ‘G’ in the same position. Sequencing of the *hbpA* gene in the 456A1 isolate according to Eriksen and co-workers (23) revealed a North Africa/C1 associated point mutation.

Multilocus sequence analysis (MLSA) of JP2 genotype isolates based on the housekeeping genes *adk*, *atpG*, *frB*, *IS 886 bp*, and *ltxCABD* (7227 bp).

**Table 1.** *Aggregatibacter actinomycetemcomitans* (A.a.) in subgingival plaque samples quantified by cultivation on blood agar plates and selective plates (TBV)

<table>
<thead>
<tr>
<th>Sample number</th>
<th>Total viable count per sample, millions</th>
<th>A.a. per sample, millions</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>455-13</td>
<td>1.2</td>
<td>0.82</td>
<td>68</td>
</tr>
<tr>
<td>456-13</td>
<td>2.4</td>
<td>1.1</td>
<td>47</td>
</tr>
</tbody>
</table>

*Fig. 1.* Schematic illustration of the leukotoxin promoter from *A. actinomycetemcomitans* genotype JP2, non-JP2, IS, and the most recent isolate 456A1. (a) The leukotoxin promoter region of *A. actinomycetemcomitans* (non-JP2): 1,109 bp (the sequence from the first nucleotide after the stop codon of glyA to the stop codon of promoter region is incorporated) (11). (b) The position of the 530-bp deletion (JP2) within the promoter region (9). (c) The position of the 886-bp insertion sequence (IS) within the promoter region (10). (d) The position of the 640-bp deletion (456 A1 isolate) within the promoter region (this study).

*Fig. 2.* *A. actinomycetemcomitans* isolates 456A1 [1], 456A2 [2], sample 455 [3], sample 456 [4], JP2 [5], and D7s [6]. (a) Strain with a complete leukotoxin promoter region, (b) strain with a 530-bp deletion in the leukotoxin promoter region, (c) strain with a 640-bp deletion in the leukotoxin promoter region. Molecular weight marker (M).
recA, and the iron acquisition–associated genes hbpA and tbpA revealed 11 sequence types among 66 isolates (24). However, none of the 11 sequence types was identical with the sequence type revealed when the 456A1 isolate was subjected to MLSA as earlier described (6). Compared with the JP2 genotype reference strain, named HK1651, a polymorphism was detected in four of the genes (Table 2). The deletion in 
\[ \text{tbpA} \]

in the 456A1 isolate has been detected only in the non-JP2 genotype of \( \text{A. actinomycetemcomitans} \) (24). This deletion may be a valuable marker for future evaluation of the dissemination routes of isolates with this mutation.

Genetic similarities between the 456A1 isolate and a JP2 genotype strain were indicated by AP-PCR (arbitrarily primed PCR) analyzes (6) (Fig. 4). This AP-PCR pattern has been detected in other JP2 genotype isolates, as well as among some of the serotype b non-JP2 genotype isolates (6).

Three different leukotoxin promoter types of \( \text{A. actinomycetemcomitans} \) have been described. In this paper, we present a promoter type which has not previously been reported. It is characterized by a missing 640-bp sequence in the leukotoxin promoter region. Like the JP2 genotype, it is highly leukotoxic according to the results obtained in leukotoxicity assays (6). In addition, it might follow a dissemination route not identical to the one for the JP2 genotype. For confirmation and further characterization of this putative novel leukotoxin promoter type, identification of additional carriers and access to additional isolates is a prerequisite.

**Table 2.** Polymorphic sites in housekeeping genes of the 456A1 isolate in comparison with the \( \text{Aggregatibacter actinomycetemcomitans} \) reference strain HK1651

<table>
<thead>
<tr>
<th>Gene</th>
<th>HK 1651</th>
<th>456A1</th>
</tr>
</thead>
<tbody>
<tr>
<td>atpG</td>
<td>231/500</td>
<td>231/500</td>
</tr>
<tr>
<td>hbpA-1</td>
<td>44/439</td>
<td>44/439</td>
</tr>
<tr>
<td>hbpA-2</td>
<td>6/329</td>
<td>6/329</td>
</tr>
<tr>
<td>tbpA</td>
<td>213/329</td>
<td>213/329</td>
</tr>
</tbody>
</table>

**Acknowledgements**

The authors thank laboratory technicians Ewa Engbo-Strömqvist and Chrissie Roth for their valuable contributions to the detection and characterization of the novel leukotoxin promoter type.
Conflict of interest and funding

The authors declare that they have no conflict of interest. This study was supported by the Research Fund (TUA) of Västerbotten County, Sweden; the Swedish Dental Society; the Danish Dental Association; and the Ingeborg and Leo Dannins Foundation.

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